



Antibody Cross-Reactivity Profiling of a Human Monoclonal Antibody with the PEPperCHIP® Human Epitome Microarray

Introduction

Antibodies are one of the most important life science tools for therapeutics, basic research and diagnostic tests. However, mono- and polyclonal antibodies are often poorly characterized in terms of specificity and cross-reactivity, as summarized in the recent Nature Feature *Reproducibility crisis: Blame it on the antibodies*:¹ The Human Protein Atlas consortium from Sweden has analyzed around 20,000 commercial research antibodies so far, and found that less than 50% can be effectively used for immunohistochemical tests. Researchers at Mount Sinai Hospital in Toronto, Canada, had been chasing a protein called CUZD1, a supposed diagnostic marker protein for pancreatic cancer. They bought a protein-detection kit comprising a CUZD1 specific antibody and wasted two years, \$500,000 and thousands of patient samples before they realized that the antibody actually recognized a different cancer protein, CA125, but did not bind to CUZD1 at all.

These two examples of the Nature Feature strongly underline the urgent need for antibody validation and cross-reactivity testing. To address this topic, PEPperPRINT has developed a three-step approach based on the new PEPperCHIP® Human Epitome Microarray² (Fig. 1).

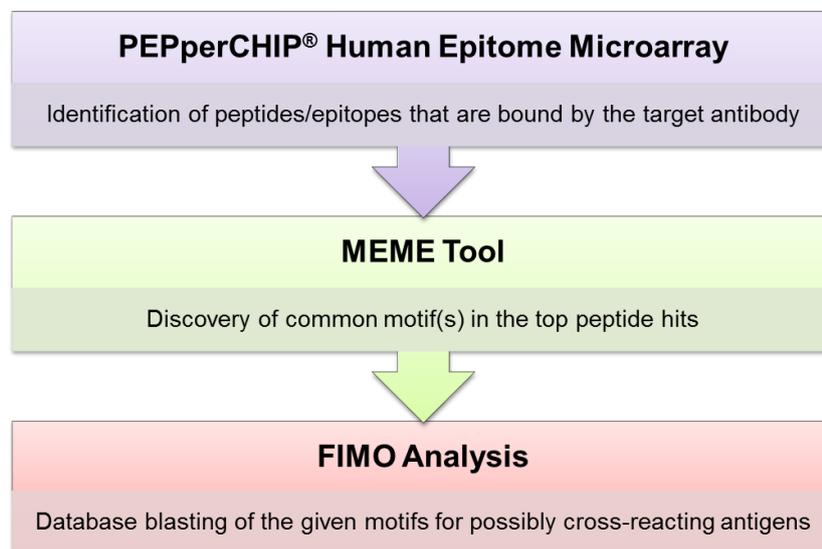


Fig. 1: Workflow of PEPperPRINT's three-step approach for antibody cross-reactivity analysis

In this application note, we describe the cross-reactivity analysis of a human monoclonal anti-c-Myc (chi9E10) antibody with the PEPperCHIP® Human Epitome Microarray and the three-step approach sketched in Fig. 1. The human monoclonal anti-c-Myc (chi9E10) antibody reacted with a number of peptides of the PEPperCHIP® Human Epitome Microarray with moderate to high signal-to-noise ratios. MEME analysis³ of the top 10 peptide hits revealed a single consensus motif xLV(S/A/P)E that was further analyzed with the FIMO tool to identify cross-reactive human antigens containing the same motif.



Results & Discussion

The PEPperCHIP[®] Human Epitome Microarray covers 29,127 linear peptides printed in duplicate (58,254 peptide spots) as well as additional polio and HA control peptides.² The peptide microarray content is based on all linear B-cell epitopes of the Immune Epitope Database⁴ with the host "human", and was further complemented by all epitopes of the most common vaccines. Therefore, the PEPperCHIP[®] Human Epitome Microarray is an ideal tool to screen for antibody responses against tens of thousands of different antigenic peptides that were described in literature and linked with antibody responses in human serum or plasma. Moreover, it enables straightforward access to scientific background information of each peptide hit by direct links to epitope entries in the Immune Epitope Database.

In a microarray pre-staining, PEPperCHIP[®] Human Epitome Microarray was initially incubated with the secondary antibody goat anti-human IgG (Fc) DyLight680 only to analyze background interactions with the printed database epitopes. Subsequently, the same microarray was incubated with human monoclonal antibody anti-c-Myc (chi9E10) with a concentration of 2 µg/ml followed by staining with the secondary antibody and control antibody mouse monoclonal anti-HA (12CA5) DyLight800. Data read-out was done with a LI-COR Odyssey Imaging System, and image analysis with PepSlide[®] Analyzer⁵.

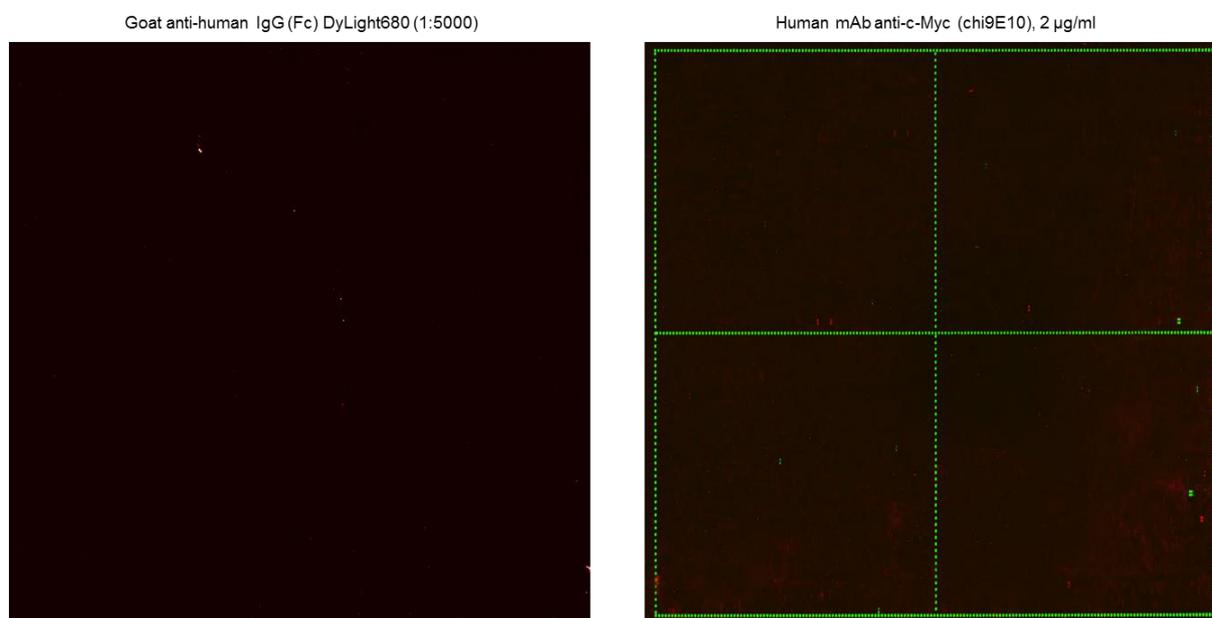


Fig. 2: Pre-staining with the PEPperCHIP[®] Human Epitome Microarray with the secondary antibody goat anti-human IgG (Fc) DyLight680 did not show any background interaction with the 29,127 database epitopes (left). Subsequent incubation of the same microarray with human monoclonal antibody anti-c-Myc (chi9E10) followed by staining with secondary and control antibodies highlighted few but clear interactions of human mAb anti-c-Myc (chi9E10) with the database epitopes in red as well as a well-defined frame of HA control peptides in green (right).

Pre-staining of the PEPperCHIP[®] Human Epitome Microarray with the secondary antibody only did not show background interactions with the database epitopes (Fig. 2, left). Incubation of the same microarray with human monoclonal antibody anti-c-Myc (chi9E10) was followed by staining with secondary and control antibodies and resulted in few but clear responses with moderate to high signal-to-noise ratios as well as a well-defined frame of HA control peptides (Fig. 2, right).



After data quantification, the top 40 interactions of human mAb anti-c-Myc (chi9E10) were sorted by decreasing spot intensities (Fig. 3). The main responses of the monoclonal antibody were directed against peptides of nucleoprotein of measles virus strain Edmonston, erythrocyte membrane-associated giant protein antigen 332 of *Plasmodium falciparum*, polymerase of hepatitis B virus and truncated Is1560 transposase of *Mycobacterium tuberculosis*. The strongest cross-reaction with a human epitope was found for dihydrolipoamide S-acetyltransferase peptides with the consensus motif LAKILVPE.

Peptide	Human mAb anti-c-Myc (chi9E10), 2 µg/ml	Epitope ID	Source Molecule Accession	Source Molecule Name	Source Organism ID	Source Organism Name
LGITAE DARLVSEIAMH	7387.7	98874	127900	Nucleoprotein	11235	Measles virus strain Edmonston
EEVVGEEKLVSEEIVT	2535,0	11927	13508497	Erythrocyte Membrane-Associated Giant Protein Antigen	5833	Plasmodium Falciparum
EVQEEGLYAKILVPE	1960,5	125443	119587578	Dihydrolipoamide S-Acetyltransferase (E2 Component Of	9606	Homo Sapiens
LAKILVPEGRDVP	1559,0	125468	119587578	Dihydrolipoamide S-Acetyltransferase (E2 Component Of	9606	Homo Sapiens
TAEDA RLVSEIAMHTTE	1050,5	98874	127900	Nucleoprotein	11235	Measles virus strain Edmonston
TYGRK LHLVSHPIILGF	1007,5	190568	4323200	Polymerase	10407	Hepatitis B virus
ALVAEGIEAIVFRTL	923,0	178535	15608176	Truncated Is1560 Transposase	1773	Mycobacterium tuberculosis
NGFLDVFTSFGGLVAE	748,0	6520	75352214	Outer Surface Protein Vlse	224326	Borrelia Burgdorferi B31
FTSFGGLVAEAFGF	639,0	18069	75352214	Outer Surface Protein Vlse	224326	Borrelia Burgdorferi B31
QYLVGERTVLAQQCYI	620,0	133331	113118	Muscarinic Acetylcholine Receptor M1	9606	Homo Sapiens
AQTQSLVYFF	593,0	78152	162797	Beta-Casein Precursor	9913	Bos Taurus
VVSVYNTNIGLKFRLQLL	547,5	53394	164509170	Precore Protein	10418	Hepatitis B virus subtype ayw
VLNPDWDQVKR	545,5	95970	115654	Alpha-S2-Casein Precursor	9913	Bos Taurus
WLSLLVFPV	496,0	72794	15211906	Large Surface Antigen	10407	Hepatitis B virus
GRSPRRRTPRRR	495,0	55713	16930336	Core Protein	10407	Hepatitis B virus
GLSPTVWLV	485,0	21139	128168864	Envelope Protein	10407	Hepatitis B virus
PSPRRRRSQSPRRR	475,5	55713	16930336	Core Protein	10407	Hepatitis B virus
WSEGEVAFYRVLDLHF	468,0	119822	62094	110 Kd Polyprotein Precursor	11041	Rubella virus
GAAATAAQAAVVRDFQ	443,0	21976	15611010	10 Kda Culture Filtrate Antigen Esxb	1773	Mycobacterium tuberculosis
PWATLVAE	418,0	140582				
KYGGTKYNGEEYU	414,0	34437	116200	10 Kda Chaperonin	1773	Mycobacterium tuberculosis
RKHLVSHPIILGFRKI	412,0	190568	4323200	Polymerase	10407	Hepatitis B virus
SSLRFG	408,0	356460				
MININIFMRESSRSFL	399,0	11711				
RRRSQSPRRR	396,5	55713	16930336	Core Protein	10407	Hepatitis B virus
HVYLDVTVLIGALAN	383,0	25128	15609123	Probable Conserved Integral Membrane Protein	83332	Mycobacterium tuberculosis H37Rv
RRRSILPYGDSMDRI	377,0	125882	146345399	Collagen Alpha-1(XVII) Chain	9606	Homo Sapiens
WWARRRRRWRWRKRR	360,0	127897	5441535	Orf1	68887	Torque Teno Virus
FPGGGQVGGVYVLP	357,0	350483	974351	Core Protein	356114	Hepatitis C Virus Genotype 3
LVAEEDER	352,5	7979	SRC279960	Genome Polyprotein	11103	Hepatitis C Virus
DVKFPGGGQI	332,0	10636	130461	Genome Polyprotein	11103	Hepatitis C Virus
GPSVFL	331,0	107421	494350	Chain H, Three-Dimensional Structure Of A Human Immunoglobulin G	9606	Homo Sapiens
AIAEYERSAAVLVRYPF	329,5	193035	564602885	Peptidylarginine Deiminase	837	Porphyromonas Gingivalis
NPGLLRFLPQLSERL	324,0	179310	15609704	Hypothetical Protein	1773	Mycobacterium tuberculosis
FLPSDFPVS	311,5	79531	116946	Capsid Protein	10418	Hepatitis B virus Subtype Ayw
ELGGKPALVPDRQVLYQ	311,0	20145	81992797	Genome Polyprotein	31647	Hepatitis C Virus Subtype 1B
VFCVQP	311,0	68440	130458	Genome Polyprotein	31647	Hepatitis C Virus Subtype 1B
QSLSFDSNPEYFDGYW	309,0	176618	226694183	Integrin Alpha-Iib	9606	Homo Sapiens
AQLLTFEAI	308,0	4002	57117045	Ppe Family Protein	83332	Mycobacterium tuberculosis H37Rv
CLLCAYSIEF	301,5	7713	124757	Ovomucoid Precursor	9031	Gallus Gallus

Fig. 3: The top 40 peptide hits of human mAb anti-c-Myc (chi9E10) on the PEPperCHIP[®] Human Epitome Microarray were sorted by decreasing spot intensities. The second column shows the fluorescence intensities of each peptide with direct links to the corresponding Immune Epitope Database entries in the third column.

The top 10 peptide hits of human mAb anti-c-Myc (chi9E10) were uploaded to the MEME tool of the MEME Suite to discover common motifs in these peptides.³ MEME represents motifs as position-dependent letter-probability matrices, which describe the probability of each possible letter at each position in the pattern. Analysis of the top 10 peptides of human mAb anti-c-Myc (chi9E10) resulted in a single consensus motif $^1xLV(S/A/P)E^5$ with a very high statistical significance of $E = 1.0e-004$ (Fig. 4, left). According to this MEME motif, amino acid 2L appeared to be essential for binding of human mAb anti-c-Myc (chi9E10), while amino acids 3V and 5E were highly conserved. Amino acid position 1 was rather variable, position 4 exhibited a clear preference for S, A and P.

The MEME consensus motif was uploaded into the FIMO tool ("Find Individual Motif Occurrences"), and processed in a protein database blast search with a focus on human proteins. The FIMO analysis resulted in 727 human proteins with the MEME motif $^1xLV(S/A/P)E^5$ or motif variants. Sorted by decreasing p-values and hence response probabilities, these proteins can be regarded as possible candidates for cross-reactions of human monoclonal antibody anti-c-Myc (chi9E10) (Fig. 4, right). The top FIMO candidates were based on the consensus motif HLVSE and assigned to DNA repair protein XRCC4, putative uncharacterized protein BVES-AS1, transmembrane protein 109 or choline O-



acetyltransferase. The expected response against human Myc proto-oncogene protein was also found among the database hits at position 175, albeit with a slightly less significant p-value.



Fig. 4: MEME analysis of the top 10 peptide hits of the PEPperCHIP[®] Human Epitome Microarray resulted in a single consensus motif ¹xLV(S/A/P)E⁵ with a very high statistical significance (left). Amino acid ²L was found to be essential for binding of human mAb anti-c-Myc (chi9E10), amino acids ³V and ⁵E were also highly conserved. Amino acid position 1 was rather variable, while position 4 exhibited a clear preference for amino acids S, A and P. FIMO analysis of this motif in Swissprot resulted in 727 human proteins as possible targets for antibody cross-reactivity sorted by decreasing response probabilities. The table shows the top 30 hits with highest statistical significances based on consensus motifs HLVSE and RLVSE (right).

Conclusion

Using our three-step cross-reactivity analysis based on the PEPperCHIP[®] Human Epitome Microarray and the MEME Suite with the MEME motif and FIMO tools, we could identify a common peptide motif ¹xLV(S/A/P)E⁵ that is recognized by test antibody anti-c-Myc (chi9E10). A database blast search with this consensus motif resulted in a probability-dependent list of human proteins as possible candidates for antibody cross-reactions. Interestingly, the c-Myc antigen was also found among the candidate proteins, albeit with a lower statistical significance than the top hits. On the PEPperCHIP[®] Human Epitome Microarray, we further identified a clear cross-reaction with dihydrolipoamide S-acetyltransferase peptides with the consensus motif LAKILVPE with the xLVPE stretch.

The results of the PEPperCHIP[®] Human Epitome Microarray and the MEME/FIMO analysis enable a cross-reactivity evaluation of a given antibody with respect to its application. A more detailed analysis of the identified common motif can be done by a PEPperMAP[®] Epitope Substitution Scan to identify tolerated amino acid exchanges and essential amino acid positions.⁶ Alternatively, selected cross-reactions can be validated by ELISA or in Western Blot analysis.

¹ <http://www.nature.com/news/reproducibility-crisis-blame-it-on-the-antibodies-1.17586>

² <http://www.pepperprint.com/products/human-epitome-microarray/>

³ <http://meme-suite.org/>

⁴ <http://www.iedb.org/>

⁵ <http://www.pepperprint.com/high-content-peptide-microarrays/>

⁶ <http://www.pepperprint.com/applications/epitope-substitution-scan/>